



Internationally indexed journal

Indexed in Chemical Abstract Services (USA), Index copernicus, Ulrichs Directory of Periodicals, Google scholar, CABI ,DOAJ , PSOAR, EBSCO , Open J gate , Proquest , SCOPUS , EMBASE ,etc.



Rapid and Easy Publishing

The "International Journal of Pharma and Bio Sciences" (IJPBS) is an international journal in English published quarterly. The aim of IJPBS is to publish peer reviewed research and review articles rapidly without delay in the developing field of pharmaceutical and biological sciences



Pharmaceutical Sciences

- Pharmaceutics
- Novel drug delivery system
- Nanotechnology
- Pharmacology
- Pharmacognosy
- Analytical chemistry
- Pharmacy practice
- Pharmacogenomics
- Polymer sciences
- Biomaterial sciences
- Medicinal chemistry
- Natural chemistry
- Biotechnology
- Pharmacoinformatics
- Biopharmaceutics



Biological Sciences

- Biochemistry
- Biotechnology
- Bioinformatics
- Cell biology
- Microbiology
- Molecular biology
- Neurobiology
- Cytology
- Pathology
- Immunobiology



*Indexed in Elsevier Bibliographic Database



For "Instruction to Authors" visit www.ijpbs.net

For any Queries, email to "editorijpbs@yahoo.in"



RESEARCH ARTICLE

BIOINFORMATICS

VIRTUAL SCREENING AND MOLECULAR DOCKING ANALYSIS FOR PREDICTING THE POTENTIAL CYCLOOXYGENASE-2 INHIBITING DRUGS IN THE TREATMENT OF CANCER**¹*EZHILARASAN .V, ¹CHINNATHAMBI. V, ²JANARTHANAN .V, ³YAZHINI K.A AND ⁴SRIDHAR .S****¹Molecular biology and Bioinformatics division, ARMATS Biotek training and research institute, ARMATS BIOTEK, Chennai, India.****²Department of chemistry, presidency college, Chennai, India****³Amirta School of Biotechnology, Amirta Viswa Vidyapeetham University, Kollam, Kerala, India****⁴Department of biotechnology, Jeppiaar engineering college, Chennai, India****EZHILARASAN .V****Molecular biology and Bioinformatics division, ARMATS Biotek training and research institute, ARMATS BIOTEK, Chennai, India.****ABSTRACT**

Cyclooxygenase-2 is being treated as one of the chief anti-cancer targets for colorectal, lung, breast, prostate and head/neck cancer. The focus of this study is to discover new ligand molecules, which can be used as a potential drug against Cyclooxygenase-2. The structure of Cyclooxygenase-2 of *Homo sapiens* was modeled using "MODELLER". The FDA approved and experimental level drugs are available in DrugBank3.0 database was screened against Cyclooxygenase-2 using the virtual screening facility offered by PYR-X0.8 software. Molecular docking studies were performed using AutoDock Wizard and the results were analyzed critically with the help of AutoDock tools 1.4.5. Virtual Screening and Molecular Docking Analysis revealed four molecules. Namely, N-cyclopropyl-4-methyl-3-[1-(2-methylphenyl) phthalazin-6-yl]benzamide, 6-Fluoro-2-(2'-Fluoro-1,1'-Biphenyl-4-Yl)-3-Methylquinoline-4-Carboxylic Acid, Eletriptan and Tamibarotene.



KEYWORDS

Cyclooxygenase-2; Anti-cancer drugs; molecular modeling and docking; virtual screening.

1. INTRODUCTION

Apoptosis is an evolutionary and conserved programme mode of cell death that is critical for the maintenance of tissue homeostasis. Also, apoptosis contributes to the cytotoxic effects of standard genotoxic chemotherapy and radiotherapy. Apoptosis signaling has been tightly regulated by two main Apoptosis pathways, which are termed as 'extrinsic' and 'intrinsic'. They involve cell surface death receptors or the mitochondria and the endoplasmic reticulum^{1, 2}. Both the pathways lead to the activation of specialized proteases; the caspases that cleave diverse cellular substrates, thereby fostering death execution. However, Apoptosis signaling pathways are disrupted or impaired in tumor cells resulting in Apoptosis resistance, which is one of the common traits that tumor cells acquire during malignant transformation³. Unfortunately, the same cellular changes that allow the tumor cells to survive micro-environmental stress during tumorigenesis can cause cross-resistance to Apoptosis induction by genotoxic therapies^{4, 5, 6}. Therefore, the current research concentrates on the identification of novel agents that induce cell death in tumor cells with resistance to Apoptosis induced by chemotherapy and radiotherapy or that enhance the efficacy of genotoxic therapies in tumor cells with Apoptosis resistance in order to improve the outcome of the treatment^{1, 2, 7}.

Cyclooxygenase-2 performs a vital role in prostaglandin biosynthesis, inflammatory cells and in the central nervous system. Prostaglandin synthesis, which is present in these sites, has a key role in the development

of inflammation and hyperalgesia⁸. COX-2 is constitutively over expressed in many human pre-malignant, malignant and metastatic epithelial tumors. Some examples include colorectal⁹, lung¹⁰, breast, prostate^{11, 12}, mammary tumors¹³, thyroid¹⁴ and ovarian cancer^{15, 16}. Up-regulated expression of COX-2 is an early event during carcinogenesis and is mostly associated with poor prognosis as it promotes tumor cell proliferation, angiogenesis, invasion and metastasis^{17, 18, 19}.

The over-expression of COX-2 in RIE cells has been shown to increase the proto-oncogene Bcl-2 and lead to inhibition of Apoptosis. The inhibition of Apoptosis, a process of cell death, appears to be a key pathway in the survival of cancer cells. Experimental models have been able to reverse this inhibition of Apoptosis by using sulindac sulfide, a nonspecific COX inhibitor. This could result in increased cancer cell death and sensitization of cancer cells to chemotherapy agents¹².

The previous studies clearly demonstrate that COX-2 involves carcinogen activation, Apoptosis inhibition, tumor invasion and angiogenesis promotion. Hence, it is reasonable to say that COX-2 inhibitors can offer an important and powerful target for cancer prevention and treatment. Nowadays, many clinical trials are done using COX-2 inhibitors in the prevention and treatment of cancer^{20, 21, 22}. Therefore, Cyclooxygenase-2 was fixed as a potential target in this study. A number of molecular docking analyses were performed throughout this study in order to list out the effective inhibitors against Cyclooxygenase-2.



2. MATERIAL AND METHODS

2.1 Sequence analysis

The protein sequences used in this project were isolated from Universal Protein Resource ²³. The templates used for homology modeling were obtained by running BLAST against the Protein Data Bank (PDB) ²⁴. On the basis of these hits given by BLAST, the required template PDB structures were downloaded from the protein data bank ²⁵ and global alignment was then performed between the COX-2 sequence and the selected template. The identities between templates were retrieved in terms of the score provided by ClustalW ²⁶.

2.2 Homology Modeling and structural analysis

The structure was modeled using effective and comparative molecular modeling software named MODELLER ²⁷. Modeled structures were then validated with the help of DOPE scores ²⁸ defined by MODELLER. Later, these structures were analyzed with the help of PROCHECK ²⁹. All the macromolecules and ligands were viewed and analyzed with the help of two molecular viewers namely Chimera ³⁰ and VMD ³¹.

2.3 Inhibitor selection and Molecular properties analysis

In order to list out effective inhibitors against Cyclooxygenase-2, modeled Cyclooxygenase-2 structures were subjected to virtual screening against all the FDA approved and experimental drugs available in Drug Bank 3.0 ³². Drug-likeness of the compounds was evaluated on the basis of Lipinski rule of 5 ³³ based on the data available in Drug Bank 3.0 Database.

2.4 Virtual screening & Molecular Docking studies

Virtual screening is a computational technique used in drug discovery research. It involves the rapid Computational assessment of large libraries of chemical structures in order to identify those structures which are most likely to bind to a drug target. PyRx is a virtual screening software for Computational Drug Discovery (CDD), which can be used to screen libraries of compounds against potential drug targets ³⁴. It uses a large body of already established open source software such as AutoDock 4 ³⁵ and AutoDock Vina. These two are used as Docking software. Python was used as a programming/scripting language. Open Babel was used for importing SDF files, removing salts and energy minimization. Finally, selected FDA approved and experimental category drugs, utilized for the purpose of virtual screening, were energy minimized using the steepest decent method ³⁶ with MMFF94 force field ³⁷.

3. RESULTS AND DISCUSSION

3.1 Molecular Modeling of Cyclooxygenase-2:

The sequence of Cyclooxygenase-2 was retrieved from Universal Protein Resource (UniProt) and its corresponding sequence id was **P35354**. It consists of 604 amino acids. This sequence was subjected to similarity search against Protein Data Bank, using the BLAST tool offered by NCBI. Later, the templates were selected on the basis of structural hits and its alignment pattern against the query sequence. The selected templates were as follows: chain A of 1PXX ³⁸, chain A of 1DDX ³⁹ and chain P of 2OYE ⁴⁰. Templates and their identity with the Cyclooxygenase-2 sequence are defined in Table 1.

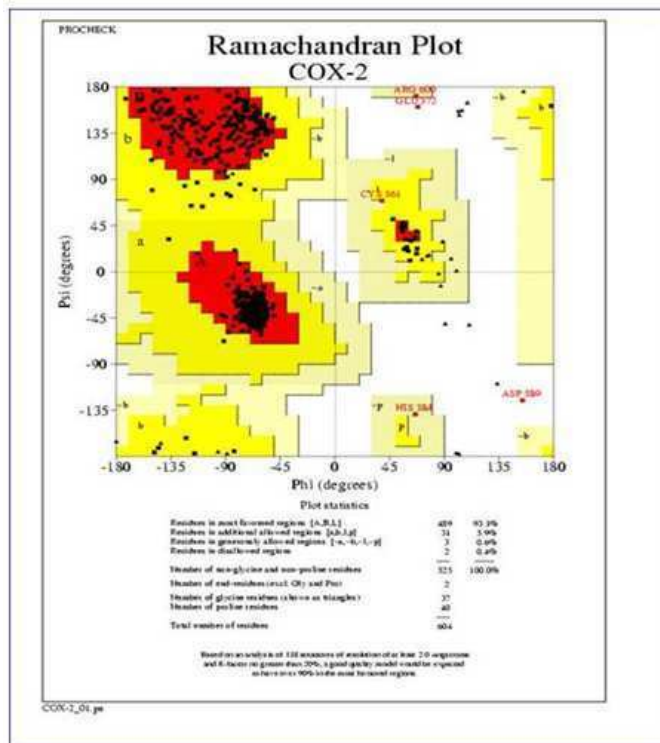
Table 1.
Templates used in Molecular modeling

S.NO	Template(PDB)	Chain	Length	Identity score with cox-2 seq
1	1PXX	A	604	86.00%
2	1DDX	A	552	88.00%
3	2OYE	P	600	59.00%

The advanced modeling tutorial package offered in MODELLER was utilized for comparative molecular modeling. Initially, Cyclooxygenase-2 sequence was converted into MODELLER input file format (.ali). Multiple sequence alignment was done using salign.py script and align_2d.py scripts and the molecular modeling was done using model-multi.py scripts. Among them, the best modeled structure was chosen with the help

of a DOPE (Discrete optimized protein energy) score. The DOPE score belonging to the best modeled structure was -69897.460938. The stereo-chemistry qualities of the structures were validated with PROCHECK structural validation tool. PROCHECK results clearly indicated the higher fidelity of modeled Cyclooxygenase-2 structure (Fig.1).

Figure 1
PROCHECK structure validation-plot





3.2 Virtual-screening

Energy minimization with universal force field ⁴¹ was done to the modeled Cyclooxygenase-2 structure using Steepest Decent method. FDA approved and experimental drugs from DrugBank3.0 were downloaded and were brought to their most stable configuration. These compounds were converted into the input-file format, namely PDBQT. During this process, drugs that had not been properly minimized and those not supported for conversion were eliminated from the list. The modeled structure was fixed as a potential target for virtual screening. Finally, virtual-screening studies were performed for all the converted drug components against modeled Cyclooxygenase-2 structure, using Vina Wizard available in PYRX-0.8 software. In the end, efficiency of all the ligands was analyzed using binding energy value predicted by PYRX-0.8 software. Binding energy is nothing but the sum of the intermolecular energy and the torsional free-energy penalty, with a more negative binding energy representing a stronger inhibition. Virtual-screening results are given in the supplementary Tables A1 and A2.

Initially, compounds described as a 'potential drug' against Cyclooxygenase-2 by UniProt Knowledgebase database were

retrieved from DrugBank 3.0. And energy minimization with MMFF94 force field was done to ligand structures using Steepest Decent algorithm. Later, compounds were subject to virtual screening and the results are described in table 2. The selected compounds and their Drugbank3.0 IDs are as follows: Acetaminophen (DB00316), Aspirin (DB00945), Balsalazide (DB01014), Bromfenac (DB00963), Carprofen (DB00821), Celecoxib (DB00482), Ciclopirox (DB01188), Diclofenac (DB00586), Diflunisal (DB00861), Epoprostenol (DB01240), Etodolac (DB00749), Etoricoxib (DB01628), Fenoprofen (DB00573), Flurbiprofen (DB00712), gamma-Homolinolenic acid (DB00154), Ibuprofen (DB01050), Icosapent (DB00159), Indomethacin (DB00328), Ketoprofen (DB01009), Ketorolac (DB00465), Lenalidomide (DB00480), Lumiracoxib (DB01283), Meclofenamic acid (DB00939), Mefenamic acid (DB00784), Meloxicam (DB00814), Mesalazine (DB00244), Nabumetone (DB00461), Naproxen (DB00788), Oxaprozin (DB00991), Phenylbutazone (DB00812), Rofecoxib (DB00533), Salicylic acid (DB00936), Salsalate (DB01399), Sulindac (DB00605), Suprofen (DB00870), Tenoxicam (DB00469), Thalidomide (DB01041), Tiaprofenic acid (DB01600), Tolmetin (DB00500) and Valdecoxib (DB00580),

TABLE 2
virtual screening of drugs currently used in medication.

S.No	Ligand	Target	Binding Energy	Molecular weight
1	DB00991	COX-2	-8.5	293.31
2	DB00482	COX-2	-8.4	381.37
3	DB01399	COX-2	-8.4	258.22
4	DB00605	COX-2	-8.3	356.41
5	DB00861	COX-2	-8.3	250.19
6	DB00939	COX-2	-8.3	296.14
7	DB00465	COX-2	-8.2	255.26



8	DB00328	COX-2	-8.1	357.78
9	DB00500	COX-2	-8	257.28
10	DB01600	COX-2	-8	260.3
11	DB00480	COX-2	-7.9	259.26
12	DB01628	COX-2	-7.9	358.84
13	DB00580	COX-2	-7.8	314.35
14	DB00784	COX-2	-7.8	241.28
15	DB01041	COX-2	-7.8	258.22
16	DB00586	COX-2	-7.7	296.14
17	DB00814	COX-2	-7.7	351.4
18	DB01240	COX-2	-7.7	352.46
19	DB00788	COX-2	-7.6	230.25
20	DB01014	COX-2	-7.6	357.31
21	DB01283	COX-2	-7.6	293.72
22	DB00533	COX-2	-7.5	314.35
23	DB00870	COX-2	-7.5	260.3
24	DB01009	COX-2	-7.5	254.28
25	DB00469	COX-2	-7.4	337.37
26	DB00812	COX-2	-7.4	308.37
27	DB00821	COX-2	-7.3	273.71
28	DB00963	COX-2	-7.3	334.16
29	DB00749	COX-2	-7.1	287.35
30	DB01050	COX-2	-7.1	206.28
31	DB00461	COX-2	-7	228.28
32	DB00712	COX-2	-6.9	244.26
33	DB00573	COX-2	-6.6	242.26
34	DB00316	COX-2	-6.5	151.16
35	DB01188	COX-2	-6.5	207.26
36	DB00936	COX-2	-6.4	138.12
37	DB00945	COX-2	-6.4	180.15
38	DB00154	COX-2	-6.3	306.48
39	DB00244	COX-2	-6.3	153.13
40	DB00159	COX-2	-6	302.45

Among these 40 compounds, Celecoxib (DB00482) and Oxaprozin (DB00991) showed significant binding energy when compared to all other compounds, these drugs are already proposed as selective Cyclooxygenase-2 inhibitors^{42, 43}. Hence, compounds exhibiting more binding energy than Celecoxib and Oxaprozin will be focused in the forthcoming analysis. Most of the drugs specific to

Cyclooxygenase-2 were reported to cause a higher risk of heart attack and stroke⁴⁴. Hence, the importance of searching new drugs against COX-2 becomes obvious.

Totally, 1480 FDA approved drugs were retrieved in the form of structural data files from the DrugBank 3.0. Among them, compounds which lacked proper coordinates were eliminated; finally 1333 compounds were subject



to virtual-screening. Likewise, 4116 compounds were selected from 5211 experimental drugs.

These 5211 experimental drugs were then filtered on the basis of physio-chemical property analysis. Small molecules having a molecular weight of more than 500 Daltons were not considered. Though a number of anti-bacterial and anti-fungal drugs showed a higher binding energy against Cyclooxygenase-2, yet, because of their higher molecular weight, they were neglected. Compounds that were directly related to the central nervous system (CNS) and steroidal drugs were also rejected, due to their high impact on other metabolic processes.

Through a detailed analysis of the characteristics of each ligand used in virtual screening, compounds having low molecular weight with better binding energy were selected from each category (approved and experimental) and considered for further study.

3.3 Molecular Docking studies:

In conclusion, selected compounds were subject to molecular docking analysis using AutoDock Module, which is available in PYRX-0.8 software. In the AutoDock Module, molecular docking was performed using Genetic algorithm parameters with a maximum of 25,00,000 energy evaluations. Later, results were analyzed with the help of Autodock tools 1.4.5. The interactions between the ligand and the target are given in figures 2 & 3. The amino acids interact with drugs are exhibiting remarkably enhanced binding affinities with Cyclooxygenase-2. The higher affinity of these small molecules is presumably attributed to the formation of hydrogen bonds. The hydrogen bond between drugs and Cyclooxygenase-2 are highlighted as green color beads (Fig.2 and fig.3).

Nearly 100 small molecules were treated in molecular docking studies, and from among them, the best compounds are listed in Table 2. Especially, Celecoxib and Oxaprozin performs a better role when compared to other small

molecules, the inhibition rate of these two compounds being highly affordable, This is obvious from the binding energy and hydrogen bonds formed between Celecoxib, Oxaprozin and the enzyme. On the other hand, there were a few compounds which exhibited a stronger inhibition when compared to the above mentioned drugs. Eletriptan (DB00216) and Tamibarotene (DB04942) among the approved category and N-cyclopropyl-4-methyl-3-[1-(2-methylphenyl), phthalazin-6-yl] benzamide (DB07307) in the experimental category showed maximum inhibition than all the other compounds. Even the supporting information associated with DB07307 in the DrugBank database stated its possible role in Apoptosis. 6-Fluoro-2-(2'-Fluoro-1,1'-Biphenyl-4-Yl)-3-Methylquinoline-4-Carboxylic acid (DB03523) from the experimental category also exhibited considerable inhibition. The interaction between the ligand and target are highlighted in figure 2, in which positions and structures of drug molecules are represented as a surface model, and the amino acids interacting with drugs are shown as a wire form model (Fig2 and 3). The predicted binding energy is listed in table 3.

Inhibitory constant represents the concentration of a drug that is required for 50% inhibition of activity of the target. The lower Inhibitory constant is more promising one for the inhibitors.

In AutoDock Inhibitory constant is calculate using following formula:

$$K_i = \exp((\Delta G * 1000.) / (Rcal * TK))$$

where ΔG is docking energy, $Rcal$ is 1.98719 and TK is 298.15

The ligand efficiency is defined as the calculated pK_i divided by the number of heavy atoms in the ligand. An affordable ligand must possess the ligand efficiency in negative. $refRMS$ is rms difference between current conformation coordinates and current reference structure. By default the input ligand is used as the reference.

Figure 2
Interaction between drugs and Cyclooxygenase-2 (FDA-Approved drug)

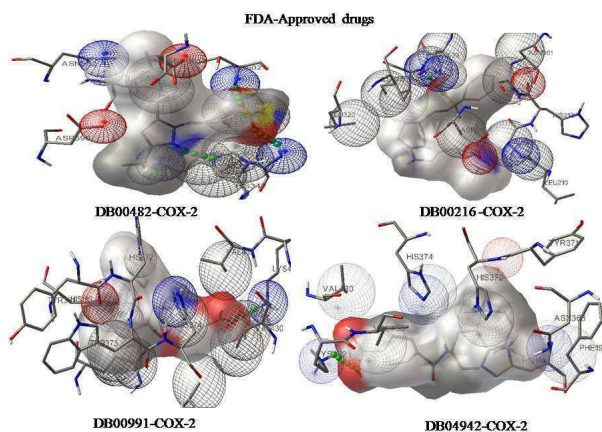


Figure 3
Interaction between drugs and Cyclooxygenase-2 (Experimental category drugs)

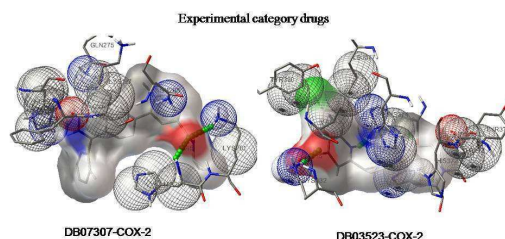


Table 3
Molecular Docking Results

S.No	Drugs	Inhibitory constant	Binding Energy	Ligand Efficiency	Ref RMS	Hydrogen Bond	Molecular Weight (daltons)
FDA-Approved drugs							
1	DB00482	8.41	-6.92	-0.27	33.73	LYS 82 THR 603 HIS 342	381.37
2	DB00991	3.6	-7.43	-0.34	37.34	LYS 432	293.31
3	DB00216	6.5	-7.08	-0.26	47.18	VAL 524	382.51
4	DB04942	1.25	-8.05	-0.31	38.05	LYS 432	351.43
Experimental drugs							
1	DB07307	2.37	-7.67	-0.26	46.12	HIS 200 LYS201	393.48
2	DB03523	1.47	-7.96	-0.28	40.97	GLN 189 LYS 432	375.36

3.4 Drug-likeness of selected compounds

The drug-likeness of the compounds was verified by a detailed analysis of the properties of drugs, chiefly properties defined in Lipinski's Rule of 5. They were as follows: Not more than 5 hydrogen bond donors; not

more than 10 hydrogen bond acceptors; and molecular weight should not be greater than 500 Daltons. The structure of the finally selected drugs is given in figure 4A and 4B. The Drug-likeness of selected compounds is given in Table 4.

Figure 4 A &4B
structure of the finally selected drugs

Figure 4 A

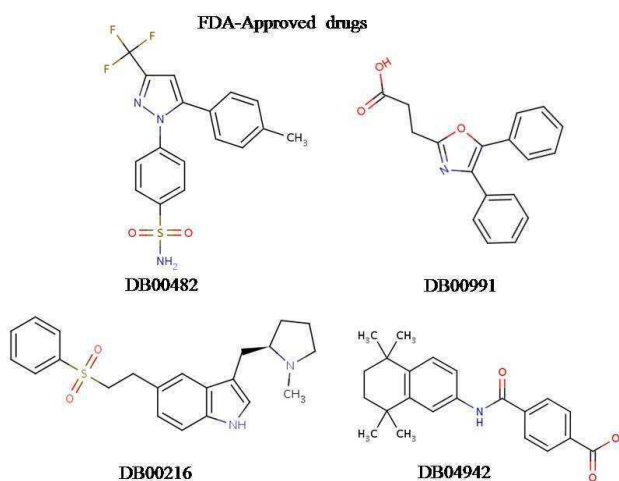


Figure 4 B

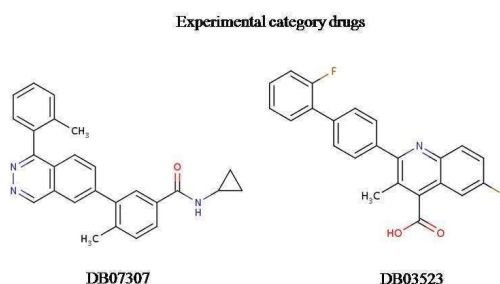




Table 4
Drug-likeness of finally selected compounds

S.No	Drugs	Log P	H-bond Acceptor	H-bond Donor	Molecular Weight (dalton)
FDA-Approved drugs					
1	DB00482	3.99	3	1	381.37
2	DB00991	3.46	3	1	293.31
3	DB00216	3.9	3	1	382.51
4	DB04942	4.99	3	2	351.43
Experimental drugs					
1	DB07307	4.77	3	1	393.48
2	DB03523	5.05	3	1	375.36

4. CONCLUSION

Virtual-screening is an emerging approach and is extensively used to reduce cost, and time in drug discovery. The approach utilized in this study was successful in searching small molecules that could act as a potential drug against Cyclooxygenase- 2 using Molecular Docking studies, compounds were screened on the basis of inhibitory constant, ligand efficiency, lowest binding energy with considerable hydrogen bonds. Hydrogen bonding plays an important

role in the structure and function of biological molecules, mainly for inhibition in a complex. Hence, the compounds N-cyclopropyl-4-methyl-3-[1-(2-methylphenyl)phthalazin-6-yl]benzamide, 6-Fluoro-2-(2'-Fluoro-1,1'-Biphenyl-4-Yl)-3-Methylquinoline-4-Carboxylic Acid , Eletriptan and Tamibarotene are strongly recommended for further clinical trials owing to their high potential to act against Cyclooxygenase-2 in the treatment of cancer.

5. REFERENCES

1. T.T. Tan, E. White. Therapeutic targeting of death pathways in cancer; mechanisms for activating cell death in cancer cells. *Adv. Exp. Med. Biol.* 615: 81-104 (2008).
2. S. Fulda, K.M. Debatin. Extrinsic versus intrinsic Apoptosis pathways in anticancer chemotherapy. *Oncogene* 25: 4798-4811 (2006).
3. D. Hanahan, R.A. Weinberg. The hallmarks of cancer, *cell* 100: 57-319 (2000).
4. T.R. Wilson, P.G. Johnston, D.B. Longely. Anti-apoptotic mechanisms of drug resistance in cancer, *Curr. Cancer Drug Target* 9: 307-319 (2009).
5. E.C. De Bruin, J.P. Medema. Apoptosis and non-apoptotic deaths in cancer development and treatment response, *Cancer Treat. Rev.* 34: 737-749 (2008).
6. M. Weinmann, V. Jendrossek, D. Guner, B. Goecke, C. Delka. Cyclic exposure to hypoxia and reoxygenation selects for tumor cells with defects in mitochondrial apoptotic pathways, *FASEB J.* 18: 1906-1908 (2004).
7. C. Belka, V. Jendrossek, M. Pruschy, S. Vink, M. Verheij, W. Budach. Apoptosis-modulating agents in combination with radiotherapy-current status and outlook,



- Int. J. Radiat. Oncol. Biol. Phys. 58: 542-554 (2004).
8. L.J. Marnett, A.S. Kalgutkar. Design of selective inhibitors of cyclooxygenase-2 as nonulcerogenic anti-inflammatory agents, *Current Opinion in Chemical Biology* 2: 482-490 (1998).
 9. M. Tsujii, S. Kawano, S. Tsuji, H. Sawaoka, M. Hori, R.N. DuBois. Cyclooxygenase regulates angiogenesis induced by colon cancer cells, *Cell* 93: 705-16 (1998).
 10. C.S. Williams, M. Mann, R.N. Dubois. The role of cyclooxygenases in inflammation, *Cancer and development, oncogene* 18: 7908-7916 (1999).
 11. S. Gupta, M. Srivastava, N. Ahmad, D.G. Bostwick, H. Mukhtar. Over-expression of cyclooxygenase-2 in human prostate adenocarcinoma, *Prostate* 42: 72-78 (2000).
 12. M. Tsujii, R.N. DuBois. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2, *Cell* 83: 493-501 (1995).
 13. R.A. Soslow, A.J. Dannenberg, D. Rush, B.M. Woerner, K.N. Khan, J. Masferrer, A.T. Koki. COX-2 is expressed in human pulmonary, colonic, and mammary tumors, *Cancer* 89: 2637-2645 (2000).
 14. V. Quidville, N. Segond, A. Tebbi, R. Cohen, A. Jullienne, M. Lepoivre, S. Lausson. Anti-tumoral effect of a celecoxib low dose on a model of human medullary thyroid cancer in nude mice, *Thyroid*. 19: 613-621 (2009).
 15. T.L. Erkinheimo, H. Lassus, P. Finne, B.P. van Rees, A. Leminen, O. Ylikorkala, C. Haglund, R. Butzow, A. Ristimäki. Elevated cyclooxygenase-2 expression is associated with altered expression of p53 and SMAD4, amplification of HER-2/neu, and poor outcome in serous ovarian carcinoma, *Clin Cancer Res.* 10: 538-545 (2004).
 16. G. Ferrandina, L. Lauriola, M.G. Distefano, G.G. Zannoni, M. Gessi, F. Legge, N. Maggiano, S. Mancuso, A. Capelli, G. Scambia, F.O. Ranelletti. Increased Cyclooxygenase-2 expression is associated with chemotherapy resistance and poor survival in cervical cancer patients, *J Clin Oncol.* 20: 973-981 (2002).
 17. J.R. Brown, R.N. Dubois. COX-2: a molecular target for colorectal cancer prevention, *J. Clin. Oncol.* 23: 2840-2855 (2005).
 18. M. Ladetto, S. Vallet, A. Trojan, M. Dell'Aquila, L. Monitillo, R. Rosato, Et al. Cyclooxygenase-2 (COX-2) is frequently expressed in multiple myeloma and is an independent predictor of poor outcome, *Blood* 105: 4784-4791 (2005).
 19. G. Ferrandina, L. Lauriola, G.F. Zannoni, A. Fagotti, F. Fanfani, F. Legge, N. Maggiano, M. Gessi, S. Mancuso, F.O. Ranelletti, G. Scambia. Increased cyclooxygenase-2 (COX-2) expression is associated with chemotherapy resistance and outcome in ovarian cancer patients, *Ann. Oncol.* 13: 1205-1211 (2002).
 20. H. Liu, Y. Yang, Y. XiaoLv, H. Yang, L. Zaho. Inhibition of cyclooxygenase-2 suppresses lymph node metastasis via VEGF-C, *Anat. Rec. (HOBOKEN)* 292: 1577-1583 (2009).
 21. D.G. Menter, R.L. Schilsky, R.N. DuBois. Cyclooxygenase-2 and cancer treatment: understanding the risk should be worth the reward, *Clin Cancer Res.* 16(5): 1384-90 (2010).
 22. S. Li, D. Tian, P. Fei, Y. Gao, Z. Chen, Q. Wang, Q. Tong. A cyclooxygenase-2 inhibitor NS-398-enhanced apoptosis of esophageal carcinoma cell EC9706 by adjusting expression of survivin and



- caspase-3, *Cancer Invest.* 29 (2): 102-6 (2011).
23. The Universal Protein Resource (UniProt). UniProt Consortium, *Nucleic Acids Res.* (2009), 37 (Database issue): D169-74 (2009).
24. S. F. Altschul, W. Gish, W. Miller, E.W. Myers, D. J. Lipman. Basic local alignment search tool, *J Mol Biol.* 215 (3): 403–410 (1990).
25. H. M. Berman, et al. The Protein Data Bank, *Nucleic Acids Res.* 28 (1): 235–242 (2000).
26. J.D. Thompson, D.G. Higgins, T.J. Gibson. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice, *Nucleic acids research*, 22: 4673-80 (1994).
27. N. Eswar, M. A. Marti-Renom, B. Webb, M.S. Madhusudhan, D. Eramian, M. Shen, U. Pieper, A. Sali. Comparative Protein Structure Modeling With MODELLER, *Current Protocols in Bioinformatics*, John Wiley & Sons, Inc., Supplement 15: 5.6.1-5.6.30 (2006).
28. D. Eramian, M.Y. Shen, D. Devos, F. Melo, A. Sali, M.A. Marti-Renom. A composite score for predicting errors in protein structure models, *Protein Science* 15: 1653–1666 (2006).
29. R.A. Laskowski, J.A. Rullmann, M.W. MacArthur, R. Kaptein, J.M. Thornton. AQUA and PROCHECK-NMR: programs for checking the quality of protein structures solved by NMR, *J Biomol NMR*, 8: 477-486 (1996).
30. H. William, D. Andrew, S. Klaus. VMD - Visual Molecular Dynamics, *Journal of Molecular Graphics*, 14: 33-38 (1996).
31. E.F. Pettersen , T.D. Goddard, C.C. Huang, G.S. Couch, D.M. Greenblatt, E.C. Meng, T.E. Ferrin. UCSF Chimera visualization system for exploratory research and analysis, *J Comput Chem.*(13): 1605-12 (2004).
32. C. Knox, V. Law, T. Jewison, P. Liu, S. Ly, A. Frolkis, A. Pon, K. Banco, C. Mak, V. Neveu, Y. Djoumbou, R. Eisner, A.C. Guo, D.S. Wishart. DrugBank 3.0: a comprehensive resource for 'omics' research on drugs: *Nucleic Acids Res. (Database issue)* D1035-41(2011).
33. C.A. Lipinski, F. Lombardo, B.W. Dominy, P.J. Feeney. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings, *Adv Drug Del Rev* 46: 3–26 (2001).
34. O. Trott , A. J. Olson. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading, *Journal of Computational Chemistry* 31: 455-461 (2010).
35. G.M. Morris, D.S. Goodsell, R.S. Halliday, R. Huey, W.E. Hart, R.K. Belew, A.J. Olson. Automated Docking Using a Lamarckian Genetic Algorithm and and Empirical Binding Free Energy Function, *J Computational Chemistry* 19: 1639-1662 (1998).
36. M.V. Fedoryuk. Method of steepest descent, in Hazewinkel, Michiel, *Encyclopaedia of Mathematics*, Springer: ISBN 978-1556080104 (2001).
37. T.A. Halgren. Merck Molecular Force Field. V. Extension of MMFF94 using Experimental Data, Additional Computational Data and Empirical Rules, *J Comp Chem* 17: 616-641 (1996).
38. S.W. Rowlinson, J.R. Kiefer, J.J. Prusakiewicz, J.L. Pawlitz, K.R. Kozak, A.S. Kalgutkar, W.C. Stallings, R.G. Kurumbail, L.J. Marnett. A novel mechanism of cyclooxygenase-2 inhibition



- involving interactions with Ser-530 and Tyr-385, *J Biol Chem*: 45763-9 (2003).
39. J.R. Kiefer, J.L. Pawlitz, K.T. Moreland, R.A. Stegeman, W.F. Hood, J.K. Gierse, A.M. Stevens, D.C. Goodwin, W. Rowlinson, L.J. SMarnett, W.C. Stallings, R.G. Kurumbail. Structural insights into the stereochemistry of the cyclooxygenase reaction, *Nature*: 97-101 (2000).
 40. C.A. Harman, M.V. Turman, K.R. Kozak, L.J. Marnett, W.L. Smith, R.M. Garavito. Structural basis of enantioselective inhibition of cyclooxygenase-1 by S-alpha-substituted indomethacin ethanolamides, *J Biol Chem*. 280: 96-105 (2007).
 41. C.J. Casewit, K.S. Colwell, W.A. Goddard III, W.M. Skiff. UFF, a Full Periodic Table Force Field for Molecular Mechanics and Molecular Dynamics Simulations by A.K. Rappe, *J Am Chem. Soc.* 114: 10024–10035 (1992).
 42. M. Farooqui, Y. Li, T. Rogers, et al. COX-2 inhibitor celecoxib prevents chronic morphine-induced promotion of angiogenesis, tumor growth, metastasis and mortality, without compromising analgesia, *Br J Cancer*. 97(11):1523-31(2007).
 43. X.P. Zhou, M.X. Zhang, W. Sun, X.H. Yang, G.S. Wang, D.Y. Sui, X.F. Yu, S.C. Qu. Design, synthesis, and in-vivo evaluation of 4,5-diaryloxazole as novel nonsteroidal anti-inflammatory drug, *Biol Pharm Bull*. 32(12): 1986-90 (2009).
 44. D. Mukherjee, S.E. Nissen, E.J. Topol. Inhibitors Risk of Cardiovascular Events Associated With Selective COX-2 Inhibitors, *JAMA*. 286(8): 954-959 (2001).